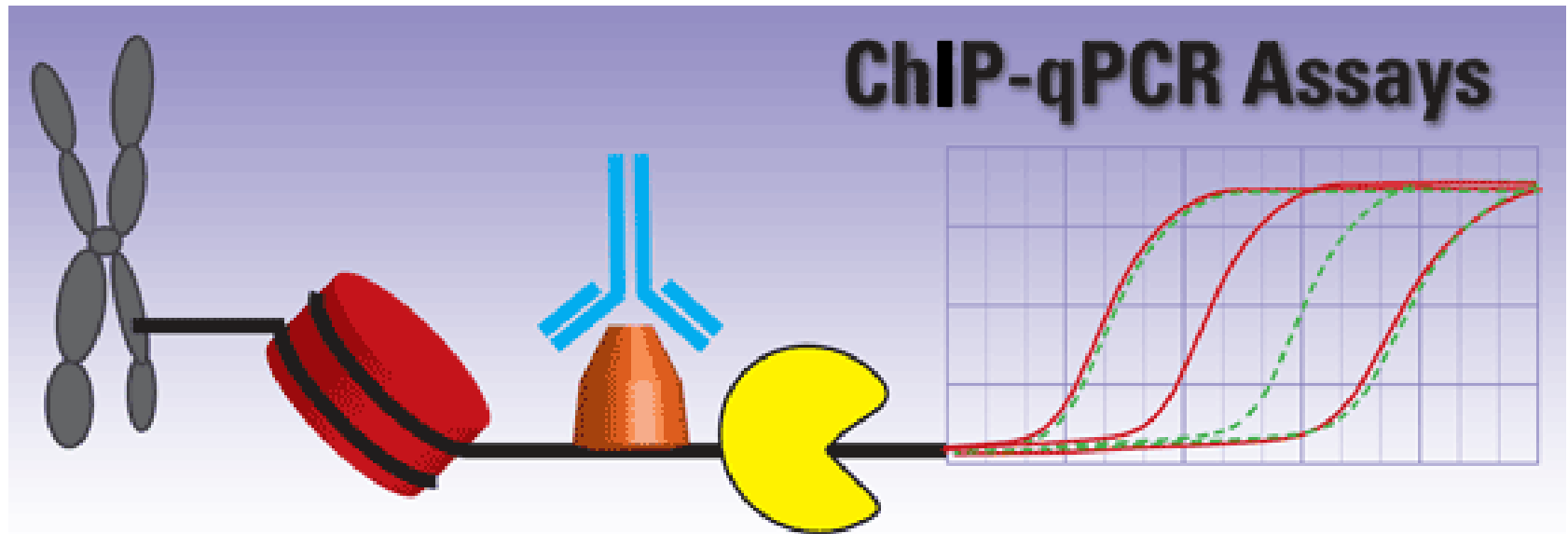


ChIP-qPCR Assays Technology Overview



**Quantitative Real-Time PCR for
Chromatin Immunoprecipitation Analysis**

“A ChIP off the old (real-time PCR) Block”



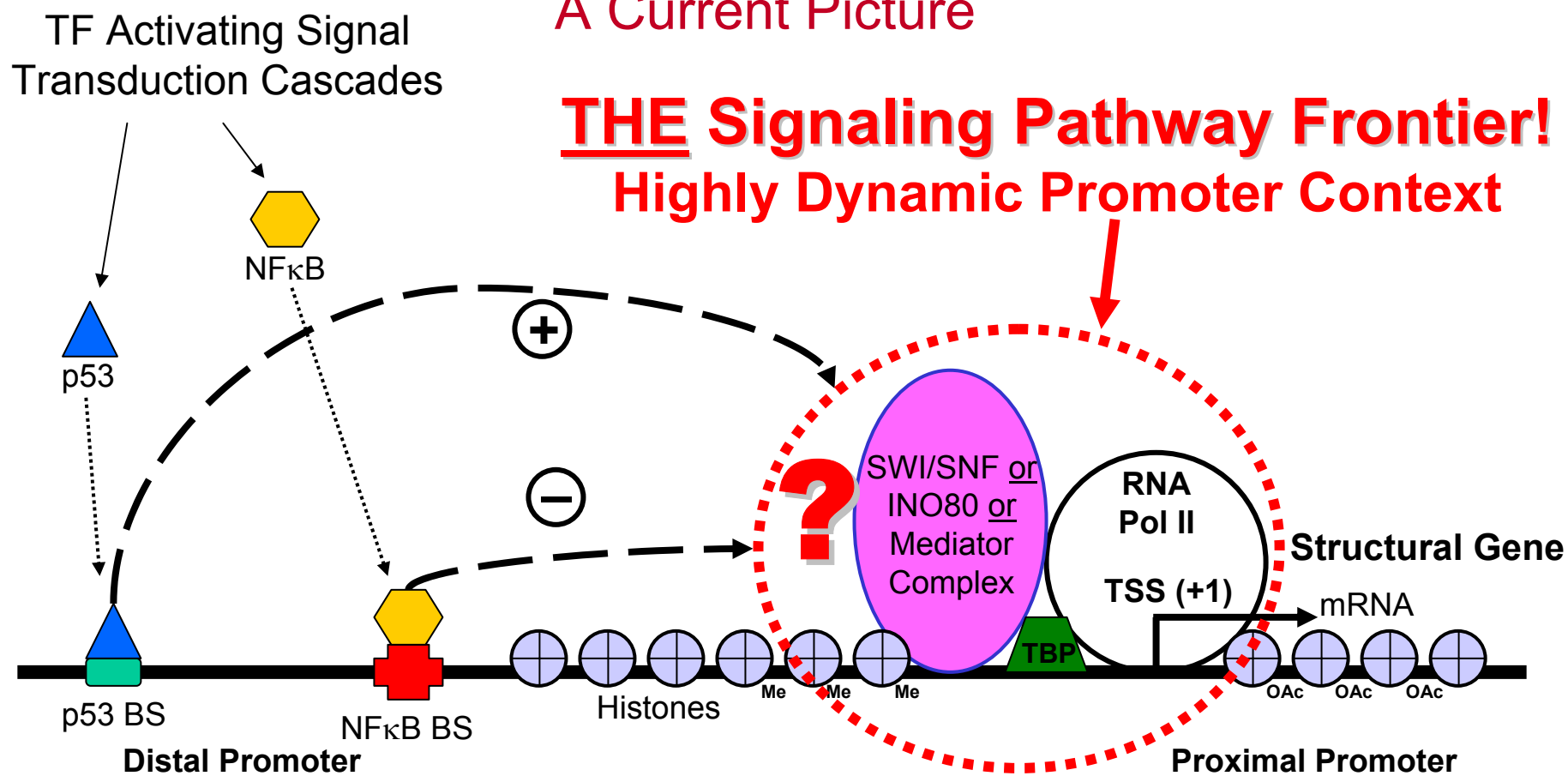
Topics to be Covered

- ❑ **Challenges Facing CHIP Researchers**
- ❑ **Solutions that the CHIP-qPCR Assays Provide**
- ❑ **How the CHIP-qPCR Assays Work**
 - ❑ Tiled Primer Assays for Every Human, Mouse, Rat Promoter
 - ❑ Quantitative Real-Time PCR Performance
 - ❑ Protocol Overview & Data Analysis
- ❑ **How YOU Can Use the CHIP-qPCR Assays**
 - ❑ Application Example:
 - ❑ p53 CHIP with CDKN1A and GADD45A qPCR

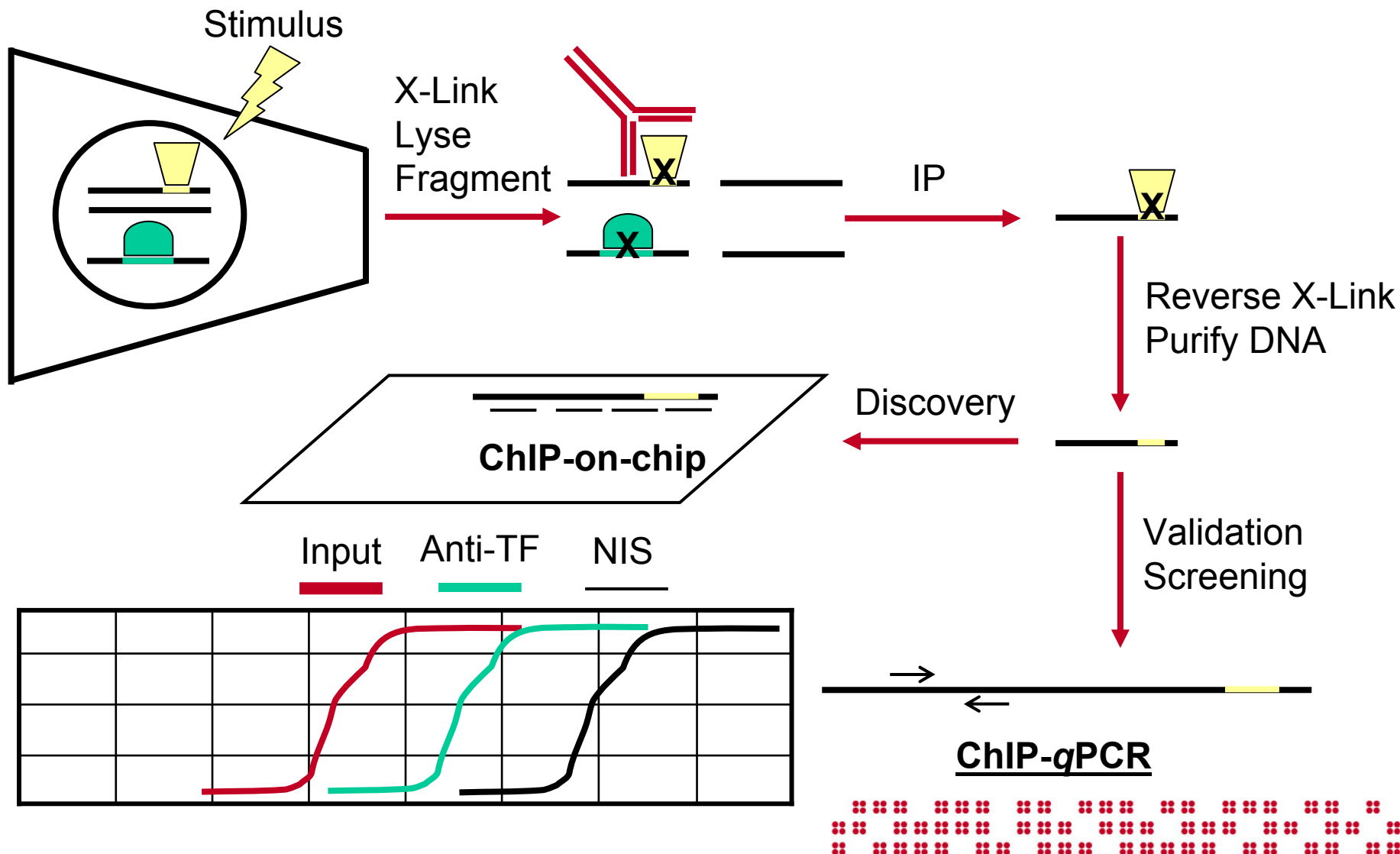


Pathway Signaling Directed Transcriptional Activation: A Current Picture

THE Signaling Pathway Frontier! **Highly Dynamic Promoter Context**



Chromatin Immunoprecipitation



Challenges Facing ChIP Researchers

- ❖ **Difficulty Designing PCR Primers for gDNA:**
 - GC-rich, high sequence content variation & repetitive sequences
 - Localizing amplicon near TF binding site can be a hindrance
 - Multiple rounds of assay design & validation
- ❖ **Time-Consuming End-Point PCR:**
 - Requires post-run gel electrophoresis & image analysis
 - NOT quantitative, narrow dynamic range, low sensitivity
- ❖ **Detecting Other Target Promoter Sites:**
 - Interesting data on one gene's promoter
 - Test another genes' promoters with SAME precious ChIP material
- ❖ **ChIP-on-chip Requires PCR Validation:**
 - Many enriched sites to validate
 - Each involving above optimization difficulties



What the ChIP-qPCR Assays Provide

⚡ Time and Money Savings:

No more repeated fruitless rounds gDNA assay design & optimization
Ready-to-use, optimized assays; Standardized PCR conditions

⚡ Quick and Easy Use:

No more post-run gel-based analysis
Quantitative & higher-throughput SYBR® Green real-time PCR
Measure enrichment at more promoter regions in same run & ChIP prep

⚡ Guaranteed Performance:

High amplification efficiency and specificity guaranteed
More accurately quantify ChIP DNA fractions, including mock IP

⚡ Comprehensive Coverage:

Tiled assays for every human, mouse or rat gene promoter
Customized genome-wide options also available

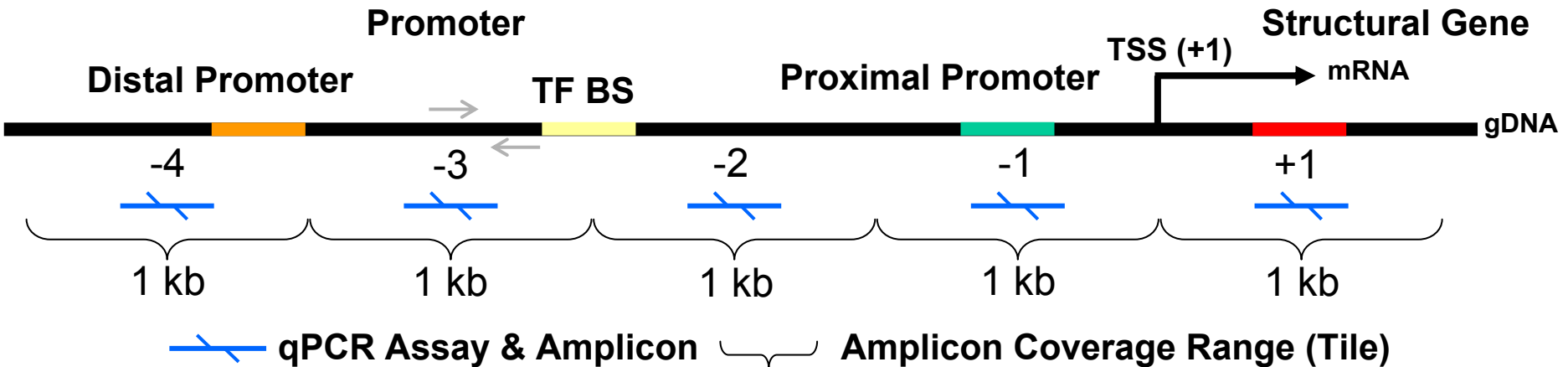


What are ChIP-qPCR Assays?

- ❖ **Pre-designed and validated real-time PCR primer assays measuring genomic DNA promoter region sequence enrichment within chromatin immunoprecipitation samples**
- ❖ Accommodate every human, mouse and rat gene promoter with a ChIP-target independent approach
- ❖ Thirty (30) 1-kb tiles from -20 kb to +10 kb relative to each TSS
- ❖ When coupled with an average 1-kb fragmentation size, provides best possible sensitivity and resolution balance
- ❖ Quality Control: One product with high amplification efficiency
- ❖ Well-suited for functional screening assays
 - ❖ Fold-differences in fractional enrichment
 - ❖ ChIP-on-chip validation



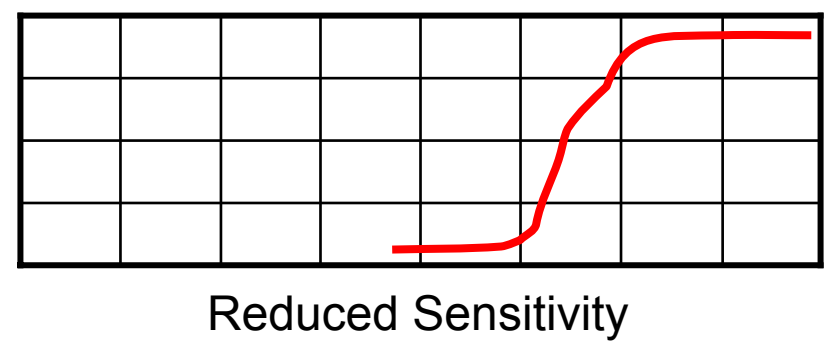
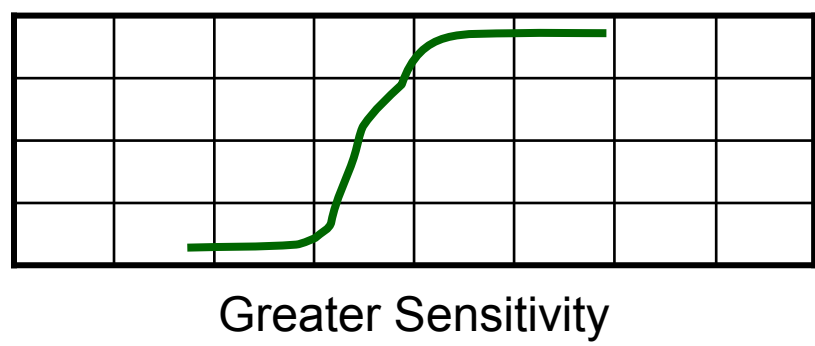
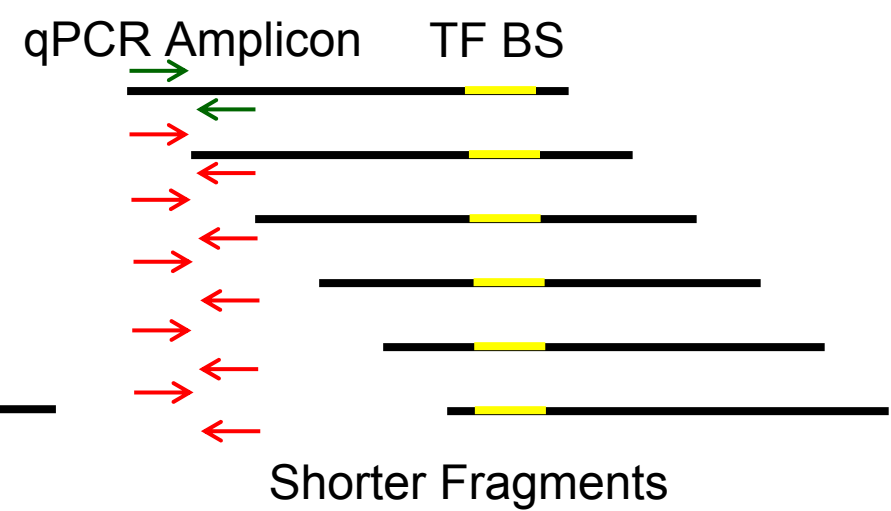
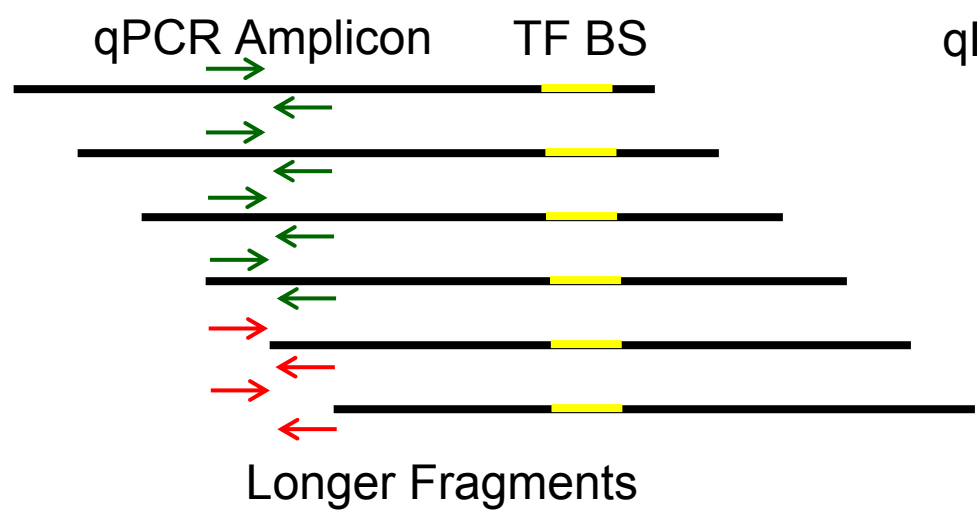
HOW IT WORKS: Tiling Scheme



- ❖ Many transcription factor binding sites (TF BS) per promoter
- ❖ Optimal qPCR primer design with stringent criteria needs room to find best amplicon sequence
- ❖ On average, amplicon is located within 0.5 to 1.0 kb of (and not necessarily at) TF binding site
- ❖ Appropriately long fragmentation size becomes critical to the success (sensitivity) of the ChIP-qPCR Assay ...



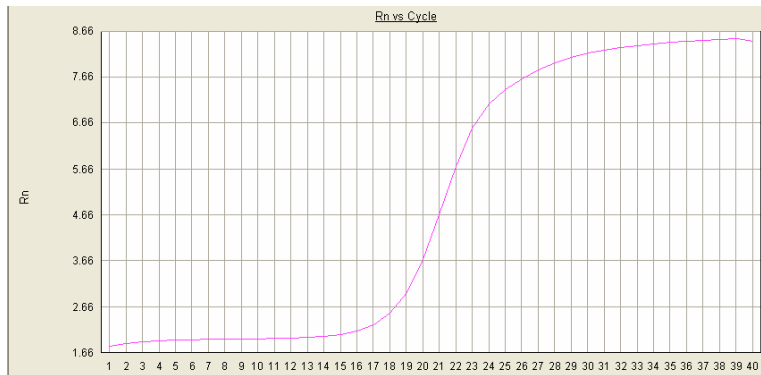
Tiling Scheme & Fragmentation Size: Resolution & Detection Sensitivity Balance



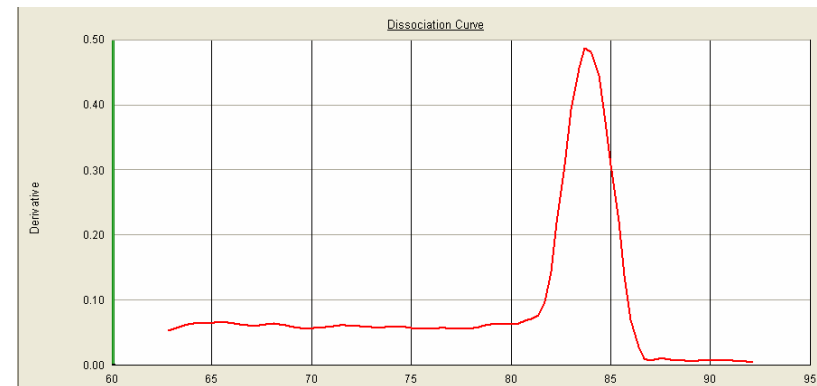
We recommend an average fragment size of 0.8 to 1.0 kb for ChIP-qPCR Assays.



Performance: Amplification Efficiency & Specificity



Consistently High Amplification Efficiencies
DART Method



Single Dissociation Curve Peak



HOW IT WORKS: Protocol Overview

- ❖ Grow & Treat Cells with Biological Replicates
- ❖ Cross-Link with Formaldehyde; Lyse Cells & Prepare Chromatin
- ❖ Fragment DNA by Sonication to **800-1,000-bp Average Size**
- ❖ **Divide Sample into Four Fractions:**
- ❖ Input gDNA (~1/100th of total)
- ❖ Immunoprecipitations:
 - ❖ Target TF = Experimental Inquiry
 - ❖ RNA Polymerase II = Positive Control for ChIP process
 - ❖ Non-Immune Serum = Negative Control for non-specific DNA IP
- ❖ Reverse Cross-Linking & Purify DNA



HOW IT WORKS: Protocol Overview

- ❖ **Three Types of qPCR Assays:**
- ❖ Promoter Target of Interest
 - ❖ Experimental Inquiry
 - ❖ **ChIP-qPCR Primer Assays, Cataloged and Custom**
- ❖ Positive Control for ChIP Process
 - ❖ Assay proximal to constitutively active promoter of an abundantly expressed housekeeping gene
 - ❖ **ChIP-qPCR Positive Control Proximal HKG Promoter Assays**
- ❖ Negative Control for Non-Specific DNA co-IP
 - ❖ Specific gDNA sequence within ORF-free intergenic region
 - ❖ “Promoter Desert”
 - ❖ **ChIP-qPCR Negative Control IGX Assays**



HOW IT WORKS: Protocol Overview

⌘ qPCR Analysis for each biological conditions' replicate:

		qPCR Assay		
		Promoter Region	Positive Control	Negative Control
ChIP Fraction	Input	√	√	√
	Anti-TF	√	X	√
	Anti-Pol II	X*	√	√
	Non-immune serum	√	√	√

⌘ Therefore, **10** reactions per biological conditions' replicate

* Unless querying proximal to promoter of interest



HOW IT WORKS: Data Analysis

ChIP	qPCR	Reports On ...
Input	Promoter Target	Total # TF Binding Sites
Anti-TF		# Binding Sites Bound by TF
Non-immune		# Binding Sites Non-Specifically co-IP
Input	HKG Proximal Promoter	Total # Pol II Binding Sites
Anti-RNA Pol II		# Binding Sites Bound by RNA Pol II
Non-immune		Background # RNA Pol II-binding sites
Input	IGX Negative Control	Amount gDNA Non-Specifically co-IP-ed with Each ChIP Antibody
Anti-TF		
Anti-RNA Pol II		
Non-immune		



HOW IT WORKS: Data Analysis

❖ Recommendations:

- ❖ Normalize each IP Fraction C_t to Input DNA Fraction C_t (ΔC_t)
 - ❖ Standardizes # sites bound by TF to total # of sites
 - ❖ Percent of input binding sites (**% Input**) = $100 * 2^{-\Delta C_t}$
- ❖ **Fold Enrichment** Above Background = $2^{-\Delta\Delta C_t}$
 - ❖ $\Delta\Delta C_t = \Delta C_t$ IP fraction - ΔC_t NIS/mock IP fraction
 - ❖ Sample specific background compensated measure
- ❖ **Differential Site Occupancy** Across Samples = $2^{-\Delta\Delta C_t}$
 - ❖ $\Delta\Delta C_t =$ Experimental IP fraction ΔC_t - Control IP fraction ΔC_t
- ❖ Positive Control for ChIP Process =
RNA Pol II **% Input** @ HKG Proximal Promoter
- ❖ Antibody Non-Specific DNA Binding Assessment =
Negative Control qPCR Assay **% Input** for each IP Fraction
matches NIS/mock IP fraction

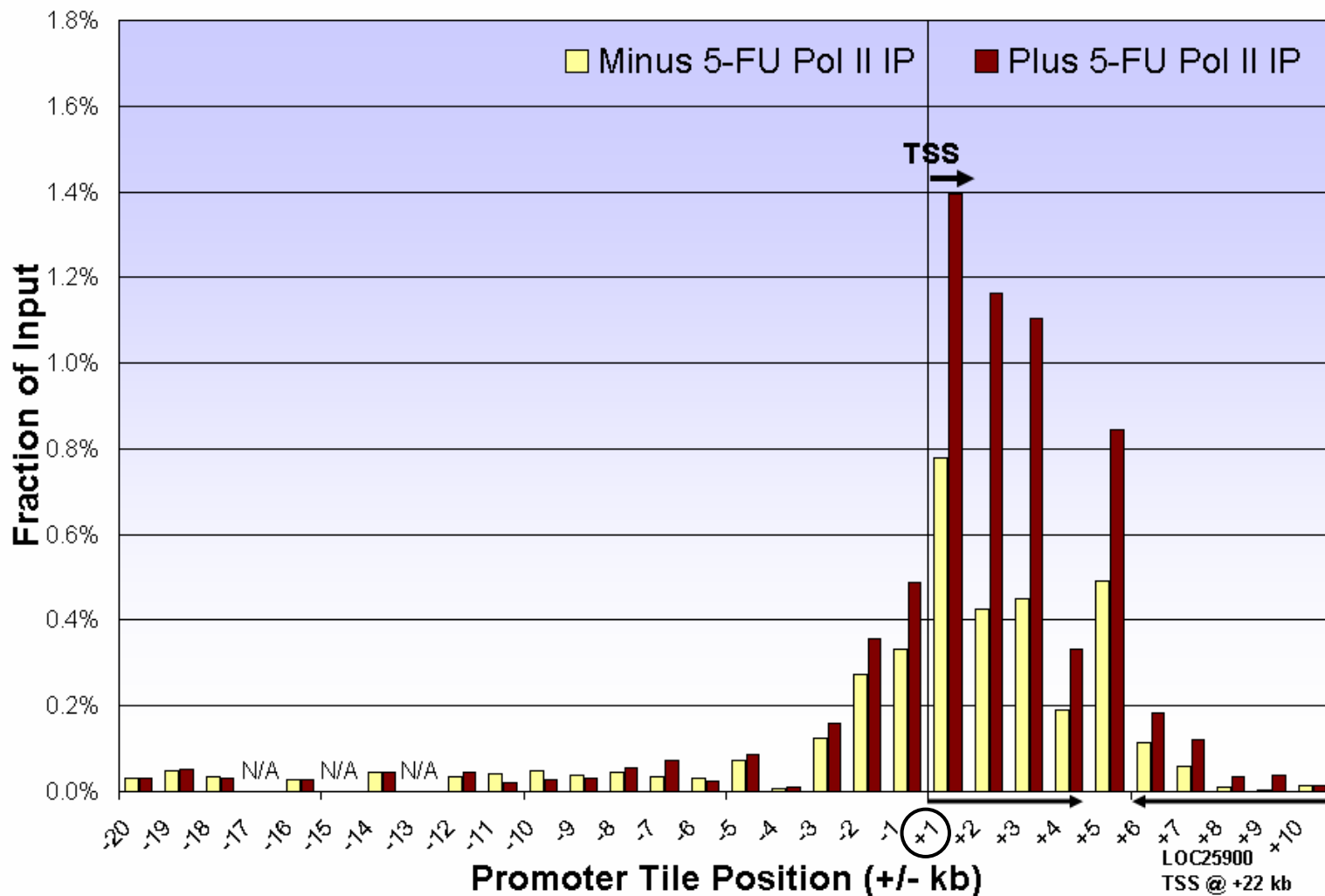


Application Example

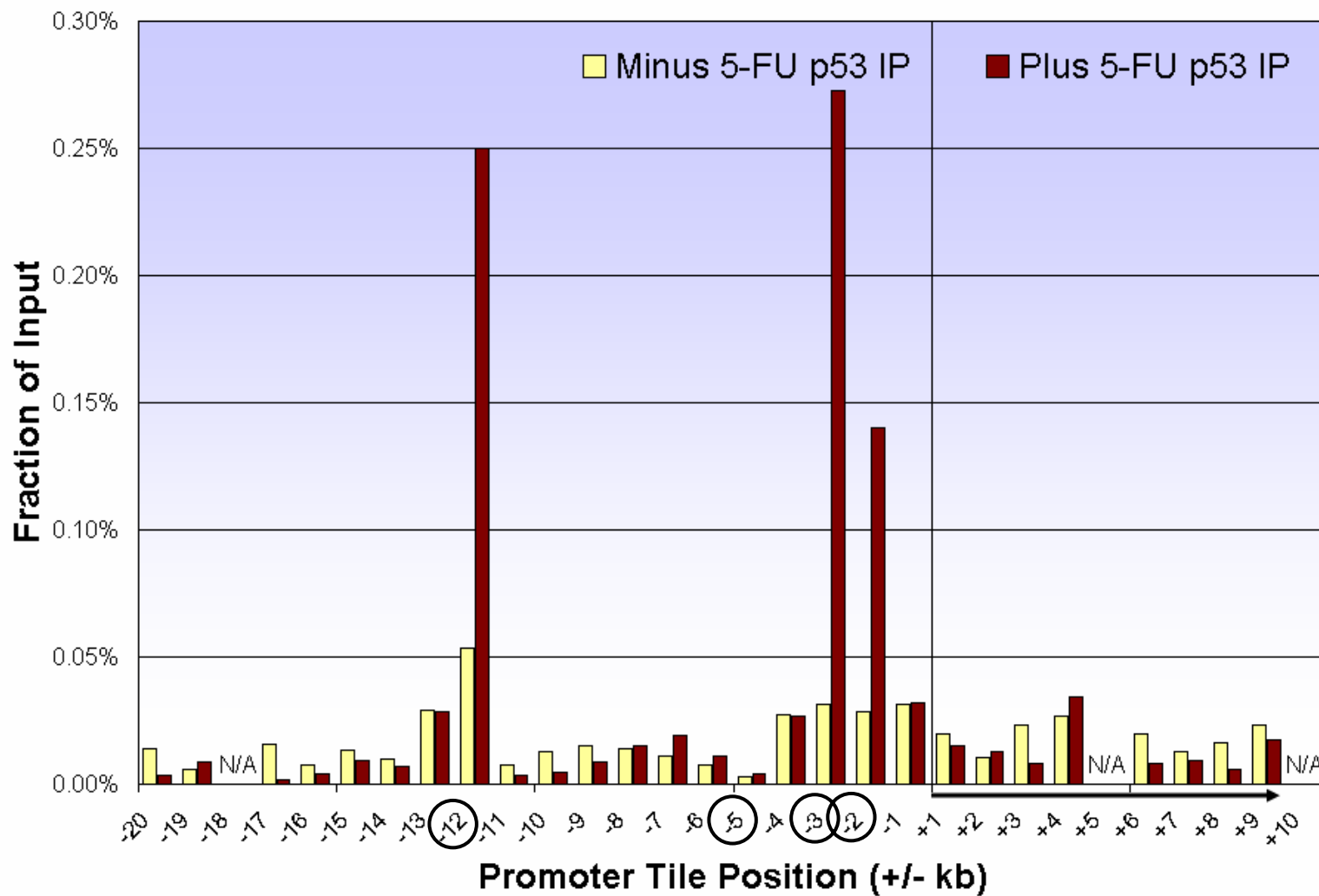
- ⌘ Treat HCT-116 cells with or without 5-flourouracil (5-FU)
- ⌘ Save fraction of total or INPUT genomic DNA
- ⌘ ChIP with anti-Pol II, anti-p53 and non-immune serum IgG (mock)
- ⌘ Measure enrichment of:
 - ⌘ p53 at CDKN1A Promoter
 - ⌘ p53 at GADD45A Promoter
 - ⌘ RNA Pol II at GAPDH Promoter
 - ⌘ IGX1A negative control region
- ⌘ Profile expression of p53 Signaling Pathway genes with RT² Profiler™ PCR Array



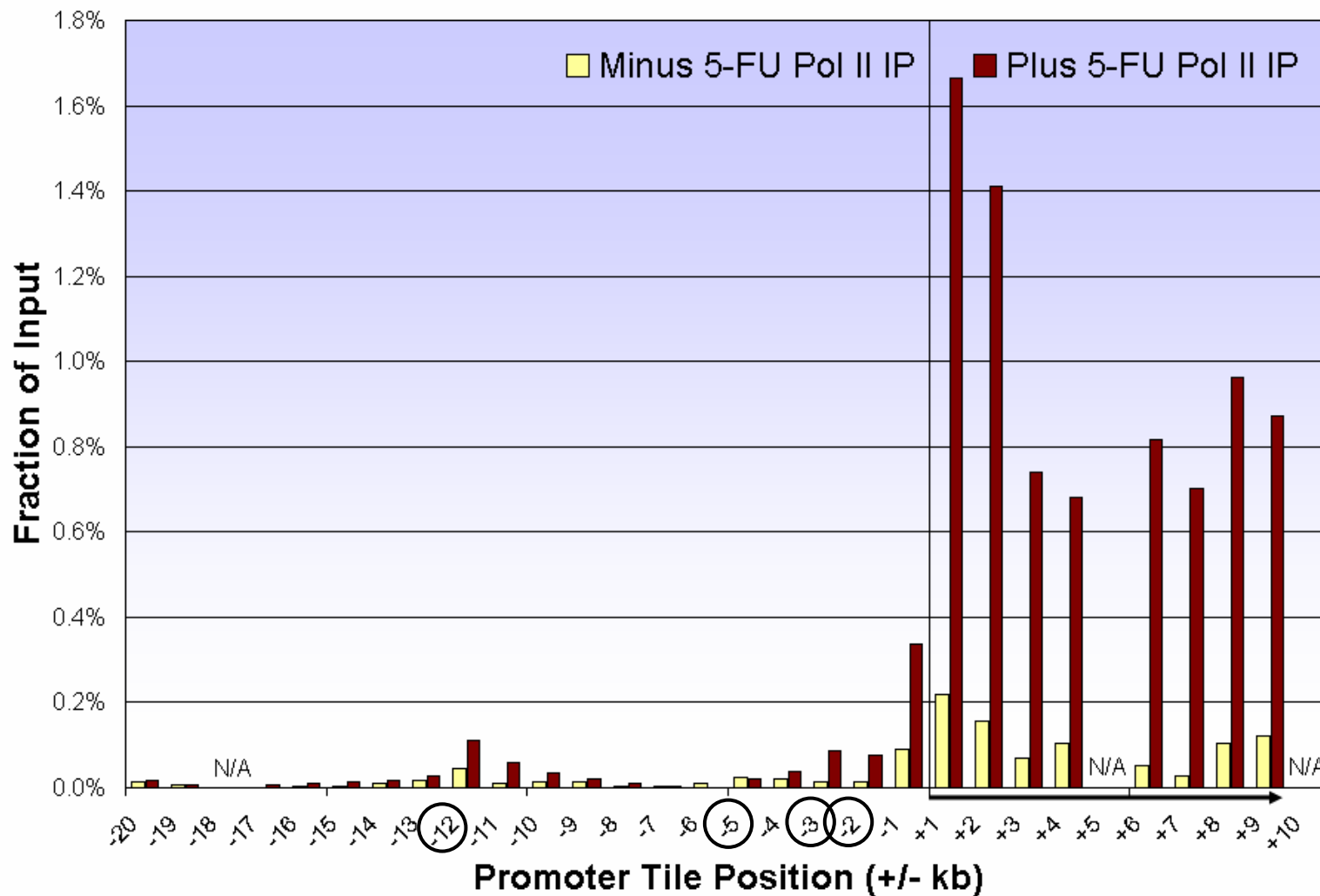
Application Example: POL II IP & GAPD qPCR Tiling



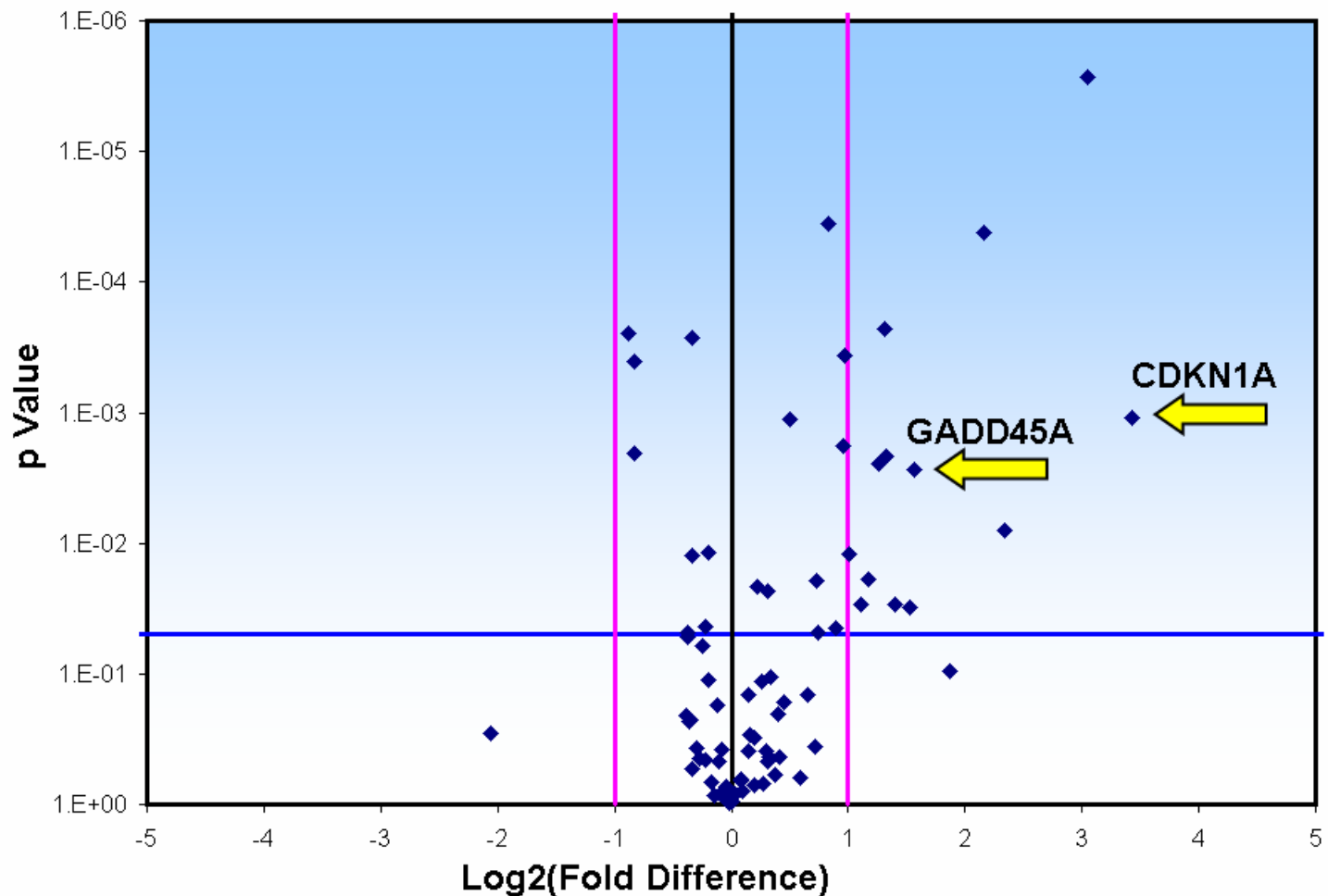
Application Example: p53 IP & CDKN1A qPCR Tiling



Application Example: Pol II IP & CDKN1A qPCR Tiling



Application Example: RT² Profiler Human p53 Signaling PCR Array



Application Example: p53 IP & GADD45A qPCR Individual TF BS Assays

ChIP Fraction	qPCR Assay	C_t	ΔC_t	Fold Enrichment	$\Delta\Delta C_t$	Fold Change
Input	GADD45A +02 Tile -5FU	29.39	-	-	-	-
RNA Pol II		33.47	10.7	0.061 %	-	-
p53		36.31	13.5	0.009 %	-	-
NIS		39.05	16.3	0.001 %		
Input	GADD45A +02 Tile +5FU	31.24	-	-	-	-
RNA Pol II		32.68	8.0	0.380 %	2.64	6.2
p53		34.06	9.4	0.146 %	4.10	17.1
NIS		38.16	13.5	0.009 %		



How to Order ChIP-qPCR Assays

- ⌘ <http://www.superarray.com/chipqpcrsearch.php>
- ⌘ Find the promoter region of interest
 - ⌘ Species then RefSeq and Tile or Chromosome Location
- ⌘ Growing set of pre-designed assays for popular TF target sites
- ⌘ Other assays designed upon request - 2 business days
- ⌘ Produced and QC on-demand with ~ one-week turnaround time
- ⌘ May be necessary to contact you if design / QC problematic to find next-best tile



Custom Assays and Plates

❖ Custom ChIP-qPCR Assays

- ❖ Whole Genome / Whole Chromosome Tiling Array Validation
- ❖ Non-promoter Regions
 - ❖ Chromatin Remodeling / Histone ChIP
 - ❖ Differentially Methylated Loci / 5-MeC ChIP, MB-PCR, MIRA
- ❖ Narrower Tiling Interval, Wider Tile Range
- ❖ *Saccharomyces cerevisiae*, *Drosophila melanogaster*

❖ Custom ChIP-qPCR Arrays

- ❖ All assays and controls conveniently arranged
- ❖ Same promoter regions, multiple ChIP experiments
- ❖ Same ChIP experiment, multiple promoter regions
 - ❖ ChIP-on-chip validation
- ❖ Promoter Tiling Experiments



SUMMARY

- ❖ **Solutions to ChIP Research Challenges:**

- ❖ **Save Time and Money:**

Don't design your own assays anymore – We can do that for you

- ❖ **Speedy and Easy Use:**

Stop running gels – Capture all of the data during one PCR run

- ❖ **Guaranteed Performance:**

Get accurate, quantitative results

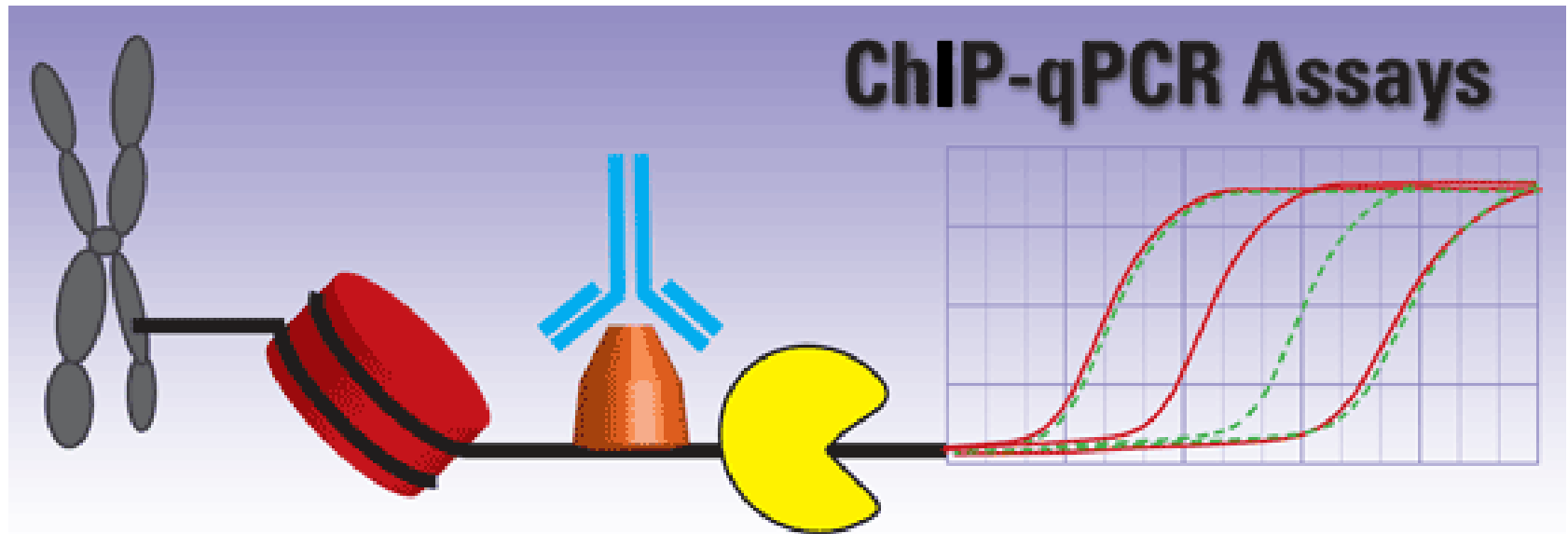
- ❖ **Comprehensive Coverage:**

Query ANY promoter or non-promoter region of interest

- ❖ **Expand your ChIP experiments to more IP targets or more promoter regions of interest with the ChIP-qPCR Assays.**



ChIP-qPCR Assays Technology Overview



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“A ChIP off the old (real-time PCR) Block”

